

Effect of different growing nutrient solutions on *Azolla pinnata* productivity under Egyptian conditions

Ahmed M. TAHA

Water Requirements and Field Irrigation Research
Department, Soils, Water and Environment Research
Institute, Agricultural Research Center, Egypt

Hamdy E. KHALIFA

Water Requirements and Field Irrigation Research
Department, Soils, Water and Environment Research
Institute, Agricultural Research Center, Egypt

A field experiment was conducted from 24 June to 31 December, 2023 in a private farm (30° 22' 01.0" N and 31° 36' 26.1" E), Egypt, to test the effects of two nutrient solutions compared with farmer practice on *Azolla pinnata* fresh biomass, tissue-chemical constituents, tissue-NPK concentrations and the amount of water applied and its water productivity. Results showed that average fresh yields were 54.9, 44.1, and 40.9 t/ha/month respectively for nutrient solution A, solution B and farmer practice. Average *Azolla pinnata* fresh yield during summer season was higher than that recorded during autumn season. In *Azolla*'s tissues, average Total Carbohydrates (TC) values were 32.3, 31.4, and 32.7%, average Total Fiber (TF) contents were 15.9, 15.7, and 15.8%, average AA values were 14.0, 12.6, and 13.1%, and average Crude Protein (CP) values were 14.0, 12.6, and 13.1% for solution A, solution B and farmer practice treatments, respectively. Average tissue-N values were 2.41, 2.21, and 2.14%, tissue-P values were 0.54, 0.40, and 0.40%, and average tissue-K values were 1.22, 1.21, and 1.25% for the same respective treatments. Total amounts of applied water during the growing period was 4071 m³/ha and average water productivity values were 96.4, 80.4, and 73.6 kg fresh yield/m³ for the three respective treatments. It could be concluded that, the multiple contents of growing nutrient solution significantly increase *Azolla pinnata* biomass. It also increased the chemical constituents of the plant, tissue-NPK, and water productivity. Ponded water contains macro- and micro-nutrients and can be used to irrigate other crops in the farm.

Keywords: *Azolla pinnata*, biomass, chemical constituents, tissue-NPK, water productivity

INTRODUCTION

Azolla is a genus of aquatic ferns and small-leaved floating plants, native to the tropics, subtropics and warm temperate regions of Africa, Asia and the Americas. It is a common free-floating fern up to 10 to 30 millimeters in diameter with roots hanging down to about 40 millimeters below the water surface. *Azolla* ferns float on the surface of the water individually or as large mats (Mosha, 2018). *Azolla* species are used as animal feed, human food and medicine, biofertilizer, water purifier, green manure, hydrogen fuel, biogas producer, weed and insect controller, and reduces ammonia volatilization after chemical nitrogen application. It improves the water quality by removing excess quantity of nitrates and phosphorus. Also, *Azolla* has many advantages and is known as a source of many essential bioactive compounds and nutrients (Ray et al., 1979; Wagner, 1997; Pabby et al., 2003; Chris et al., 2011; Sadeghi et al., 2013; Selvaraj et al., 2014; Rashad, 2021).

Few research reported the water consumption by *Azolla* plants through the growing season. The experiments by Amro (2022) showed that the monthly water consumption rate by the *Azolla* plant is

approximately equal to 120 m³/dunum (1200 m³/ha/month).

Azolla plants support nitrogen fixing bacterium, which allows it to use nitrogen from the water and air for its own growth. It can fix atmospheric nitrogen due to the presence of blue-green algae (*Anabaena azollae*) located in cavities of the ferns' lobes (Adzman et al., 2022). It is a good source of protein and contains almost all essential amino acids and minerals like iron, calcium, magnesium, potassium, phosphorus and manganese (Brouwer et al., 2018; Patil and Patil, 2020).

Under optimum growth conditions, including water depth, nutrient concentration, pH, relative humidity, air and water temperatures, and sunlight exposure, Azolla spreads across water surface until it covers the whole surface of the water in a dense cover. Azolla can double its leaf area in seven days if conditions of high nutrient levels and water temperatures persist (Adzman et al., 2022). Results by Chatterjee et al. (2013) showed that, nutrient composition of Azolla species varied depending on the environmental conditions, including temperature, light intensity, and soil nutrients. These factors would therefore have an impact on growth morphology and its nutrient composition.

Watanabe et al. (1977) conducted laboratory studies which showed that Azolla, grown in a nitrogen free solution, can double its mass in 3-5 days and can accumulate 30-40 kg N ha⁻¹ in two weeks. Results by Kannaiyan et al. (1981) and Kannaiyan (1982) indicated that P, K, Ca, Mg, Fe, Mo, Co, and Zn have been shown to be essential for Azolla growth and N-fixation. The main macronutrients and other essential nutrients that are necessary for optimizing Azolla growth and N fixation are P, K, Ca, Mg, Fe, Mo, Co, and Zn (Carithers et al., 1979; O'Hara, 2001; Kannaiyan, 1982). Previous studies showed that, Ca and P deficiencies had a considerable effect on Azolla growth and N fixation compared to K and Mg deficiencies (Watanabe et al., 1977; Subudhi and Singh, 1978; Kannaiyan et al., 1981). As the P level drops in the growth medium, it will affect growth rate and N fixation.

Also, Azolla growth was reduced in low concentrations of Fe, Ca, or P. Nordiah et al. (2012) stated that, Azolla expands its population depending on the availabilities and contents of nutrients in the growing media, while water without phosphate showed low Azolla growth. Hossain et al. (2021) concluded that, phosphorus content of Azolla pinnata was proportional to the phosphorus supplementation in the culture medium. The supplementation of 10 ppm phosphorus to water used for culturing Azolla pinnata is optimum. It also improved the protein and lipid contents of Azolla pinnata under outdoor conditions.

Results by Adzman et al. (2022) indicated that Azolla growth was the best in water depth of 20 cm, the nutrient concentration of 812.5 ppm, pH of 7 and under 100% sunlight exposure. It can survive within a water pH range of 3.5 to 10, but optimum growth occurs in the pH range of 4.5 to 7 and temperature range of 18°C to 26°C.

The tested hypothesis here is that, the combined solutions of essential nutrients improve Azolla's yield and increase nutrient concentrations in the Azolla plant tissue. Therefore, the overall objectives of the implemented field experiment were to:

- Test the effects of different nutrient solutions on the productivity of Azolla pinnata under field conditions;
- Test the effects of nutrient solutions in the Azolla growing medium on total carbohydrates, total fiber, total amino acids, and crude protein as well as on NPK nutrient concentrations in the Azolla plant tissue.
- Determine the amount of water required to grow Azolla pinnata since very few data were reported on water used by Azolla plants and on its water productivity.

MATERIALS AND METHODS

Experimental site

A field experiment on the response of *Azolla pinnata* to different nutrients fertilization was carried out for 190 days (24 Jun. – 31 Dec., 2023) in a private farm (30° 22' 01.0" N and 31° 36' 26.1" E), Sharqia governorate, Egypt (Figure 1). Based on Köppen-Geiger classification, the climate of the site is arid desert-hot (BWh) of the Mediterranean type, with most of the rainfall occurring in the winter season (Beck et al., 2018). The main daily weather data (<https://power.larc.nasa.gov/data-access-viewer/>), including maximum and minimum air temperature (°C), relative humidity (%), wind speed (m/s) and rainfall (mm), characterizing the experimental site is illustrated in Figure 2. Annual rainfall at the experimental farm from January to December 2023 was 96.1 mm, while it was 25.7 mm during the growing period (24 June to 31 December, 2023). Average maximum temperature ranged from 41.6 °C (Jul.) to 24.1 °C (Dec.), while average minimum temperature varied from 23.4 °C (Aug.) to 13.4 °C (Dec.). The mean relative humidity values varied from 43.2% (Jul.) and 67.2% (Dec.). The obtained weather data were used to calculate reference evapotranspiration (ET_o) values by applying the FAO-56 Penman-Monteith equation (Allen et al., 1998) in the FAO-CROPWAT 8 model.

Effective rainfall

The effective rainfall during the growing period was calculated on daily basis according the following relation (Dastane, 1974):

Where:

Re: effective rainfall (mm), P: depth of rainfall (mm)

Experimental design

A randomized complete blocks design (RCBD) with three replicates was used to conduct the field experiment. Three treatments, including two different combined nutrient solutions and farmer practice, were tested in this experiment.

Tested variables

The constituents of the two nutrient solutions and farmer fertilization practice tested in the experiment are:

- Solution A: Calcium Chloride (CaCl₂·2H₂O), Calcium Nitrate (Ca(NO₃)₂·6H₂O), Potassium Phosphate Monobasic (KH₂PO₄), Magnesium Sulfate Heptahydrate (MgSO₄·7H₂O), Phosphoric acid (80%), Boric acid (H₃BO₃, Boron 13%), Fe (EDTA, 13%), Zn (EDTA, 13%), and Mn (EDTA, 13%).
- Solution B: Calcium Nitrate (Ca (NO₃)₂·6H₂O), Phosphoric acid (80%, 50% of A), Boric acid (H₃BO₃, Boron 13%), Fe (EDTA, 13%), Mn (EDTA, 13%), MnSO₄·7H₂O, CuSO₄, Na₂MoO₄·2H₂O, and Manure.
- Farmer practice: Calcium Super Phosphate (15.5%), Manure, and Foliar spray of some macro- and micro-nutrients.

Cultural practices for growing *Azolla pinnata* (var. *pinnata* R. Brown)

Nine rectangular open-top aquaculture earth ponds (27.6 m x 2.9 m x 0.35 m) were prepared at the

private farm to grow *Azolla pinnata*. The earth ponds were covered with plastic sheets (high density black PE, 50 μm) to prevent water seepage. The plastic sheets were covered with a thin (10 – 15 cm) soil layer. The analysis of the soil used to fix the plastic sheet was done according to Tan (1996) and the obtained values are given in Table 1. All sides of the ponds were secured properly by placing bricks over the side walls.

The field experiment started on June 24, 2023 and each pond was filled with 18 m³ of fresh water for *Azolla pinnata* propagation. The analysis of water (Tan, 1996) used to fill the ponds is given in Table 2. The amounts of water applied were measured by flow meters. Water was applied to the ponds every two weeks and maintained at water levels between 20 and 22 cm for proper propagation.

The ponds were inoculated with *Azolla pinnata* (*pinnata* variety) at the rate of 40 kg fresh ferns/pond (0.5 kg/m²). There was a subsequent harvesting of *Azolla* at weekly interval during the period from July to October and every two weeks during November and December due to the unsuitable weather conditions for *Azolla* propagation (high wind speed and low temperatures).

Fertilization

The main fertilizers used in conducting the field experiments, element concentrations, doses, and dates of applications are presented in Table 3. For nutrient solutions A and B, macro- and micro-nutrients were added by the venturi fertilizer injector while filling the ponds with water. The farmer applied fertilizers by broadcasting and foliar methods. In solution B and farmer practice ponds, 0.5 m³/pond of manure was added. Manure analysis is given in Table 4.

Pest control

Two insecticides were used to control the pests (mainly larva) affecting *Azolla*. The two insecticides were Lambda cyhalothrin (5%) and Clorzone (48%). These insecticides were used once a week at the rate 10 cm³/20 liters of water. In case of infection and changing *Azolla* plants from green to brown color, the insecticides were used 3 times/week.

Data collection and measured parameters

The parameters measured in this study were the amounts of water applied during the growing period using water flow meter and the fresh biomass. Plant samples were collected and analyzed for total carbohydrate (%), total fiber (%), total amino acids (%), protein content (%) and NPK macro-nutrients. The chemical composition of *Azolla* was analyzed according to AOAC (2016). Also, water samples from the ponds were collected for EC, pH, cations, anions, N-NH₄, N-NO₃, and phosphorus and B, Cu, Fe, Zn, and Mn micro-nutrient analysis.

Statistical analysis

All obtained data were statistically analyzed using the MSTAT-C computer software package. For determining the effect of growing months and fertilizer treatments on the fresh yield, a two-way analysis of variance (ANOVA) was performed. The least significant difference (LSD) method was used to test the differences between treatment means at the 5% level of probability as described by Snedecor and Cochran (1981).

RESULTS AND DISCUSSION

Effect of tested treatments on *Azolla pinnata* fresh biomass

There were significant effects of the nutrient solutions used as well as the growing period on the

fresh yields of *Azolla pinnata* (Table 5). The fresh yields, although fluctuated during the growing period, were consistently higher in A-solution treatment compared to those obtained from the other two treatments. Applying nutrient solution A recorded the highest average fresh yield (54.9 t/ha, 1.71 t/ha/day), followed by nutrient solution B (44.1 t/ha, 1.39 t/ha/day), while the lowest average yield (40.8 t/ha, 1.28 t/ha/day) was obtained from farmer practice (Table 5). The obtained results were close to those reported by Abdull Aziz (2012) who indicated that the fresh yield of *Azolla pinnata* varied from 820 to 1220 kg/ha/day. The results were also similar to those of Hossain et al. (2021), who showed that the fresh biomass of *Azolla pinnata* varied from 3.90 to 5.92 kg/m² and close to those reported by Amro (2022), who indicated that the fresh yield of *Azolla* plant was equivalent to 6-7 kg/m². Results indicated also that, nutrient solution (A) that included multiple nutrients (i.e. Ca, K, P, Mg, B, Fe, Zn and Mn) significantly increased *Azolla pinnata* fresh yield by 24.5 and 34.5% as compared with the fresh yields of solution B and farmer practice, respectively. The obtained results were similar to those reported by Kannaiyan (1982) and Sadeghi et al. (2013), who showed that *Azolla* requires all macro and micro nutrients (i.e. P, K, Ca, Mg, Fe, Mo, Co, and Zn) which are essential for *Azolla* growth and N-fixation. Results agreed also with those of Nordiah et al. (2012), who indicated that a combination of more than one nutrient or multiple nutrient contents explained the increase in biomass of *Azolla pinnata*.

Results showed significant decrease in the fresh yields obtained during September as compared with those recorded in August and October. Yield reduction was due to severe shortage of the fresh water diverted to the experimental farm with direct effect on water depths in the ponds which decreased by 2 to 3 cm. The obtained results could be supported by those of Biswas et al. (2005) and Sadeghi et al. (2012a,b), who concluded that the optimal growth and biomass production of *Azolla* could have a close relation to water depth since low water depths might slow down the growth and reduce its biomass production. The obtained results could be explained by what was reported by Adzman et al. (2022) who stated that, higher volume of water could hold more dissolved oxygen (DO) providing the roots of *Azolla* with sufficient DO than lower volume of water. Also, water at deeper level is colder compared to at the surface which is exposed to direct sunlight making it warmer than water at deeper level. The cold water hold more DO because water molecules are closely packed together making it difficult to release into the atmosphere and the solubility of oxygen is decreased with warm water making it holding lesser DO.

Results showed also that, average *Azolla pinnata* fresh yield of 59.7 t/ha obtained during summer season (Jul. and Aug.) was higher than the average yield of 42.2 t/ha recorded during autumn season (Sep., Oct. and Nov.). The highest average fresh yield of 62.5 t/ha occurred during July, while the lowest average yield (22.9 t/ha) was recorded during November. The optimum weather conditions during summer, including air temperature, relative humidity, and wind speed, resulted in higher yield compared to the yield during autumn and winter seasons (Figure 3). The lowest yields during November were due to high wind speed. The obtained results were in line with those reported by Amro (2022), who found that growth rates of *Azolla* were higher during the summer. Abdul Aziz (2012) reported low productivity during winter and spring was attributed to low humidity, light intensity, day length and temperature. Also, Sadeghi et al. (2013) stated that wind and turbulent water can fragment and kill *Azolla*.

From the obtained results it could be concluded that, applying nutrient solution (A) that include multiple nutrients (i.e. Ca, K, P, Mg, B, Fe, Zn and Mn) by injecting it to the ponded water using vensuri during summer season gives the highest yield of *Azolla pinnata*.

Effect of tested treatments on total carbohydrates (TC), total fiber (TF), total amino acids (AA), crude protein (CP) in the *Azolla pinnata* tissues

Results illustrated in figure 4 showed no effect of the tested nutrient solutions on total carbohydrates (TC) values measured in *Azolla*'s tissues. Average TC values were 32.9, 31.7, and 32.9% for solution A, solution B and farmer practice treatments, respectively. The obtained results were close to carbohydrate contents reported by Mohamed et al. (2018) with 30.5% on dry matter

basis.

Meanwhile, average total fiber (TF) contents in Azolla's tissues were 16.0, 15.6, and 15.6% for solution A, solution B and farmer practice treatments, respectively. The results of crude fiber content obtained in Azolla pinnata tissues were slightly higher than the values reported by Chatterjee et al. (2013) with 13.4%, Kumar et al. (2018) with 11.2%, Wagh et al. (2021) with 14.7%, and Yee et al. (2022) with 12.2%. The slight difference between the result obtained and previous studies in the crude fiber values may be due to a change in dry matter content and maturity level of the Azolla that was collected at different intervals (Bhatt et al., 2020).

Results in figure 4 showed also that, average values of total amino acids (AA) were 14.0, 12.6, and 13.1% for the three respective treatments. For A nutrient solution treatment, the AA contents in Azolla's tissues were 11.5 and 6.9% higher than the AA contents in B nutrient solution and farmer practice treatment, respectively. The obtained results were higher than the values reported by Mohan et al. (2020), who showed that Azolla has 7-10% amino acids.

Results showed also that, average crude protein (CP) values were 15.7, 13.5, and 13.4% for the three respective treatments. In Azolla's tissues, the crude protein values obtained from A nutrient solution treatment were 16.5 and 17.6% higher than the values obtained from B nutrient solution and farmer practice treatments, respectively. The obtained results were less than the reported crude protein values from other studies showing the CP% values of 17.6% (Van Hove and Lopez, 1987), 21.2 % (Sujatha et al., 2013), 21.4% (Alalade and Lyayi 2006), 21.7 %, (Kavya 2014), 22.5 % (Ashraf and Sharma 2015), (Brouwer et al., 2018), 26.5% (Bhatt et al., 2020), 28.5% (Hossain et al., 2021), 24.1% (Yee et al., 2022), 27.1% (Adzman et al., 2022), and 25-30% (El-Naggar and El-Mesery, 2022). The low protein values could be related to the high maximum temperatures that exceeded 35 oC for several months at the experimental site with direct effect on N-fixation. The obtained results were explained by Bhatt et al. (2020), who related the variation in crude protein percentage to several conditions including air and water temperatures, nutritional content of the water, and pest growth that may affect Azolla pinnata growth and composition.

Effect of tested treatments tissue-NPK of Azolla pinnata

The effect of tested treatments on nitrogen (N, %), phosphorus (P, %), and potassium (K, %) in Azolla pinnata's tissues is illustrated in Figure 5. Results indicated that tissue-N contents (%) in Azolla pinnata varied between 3.1 – 2.1%, 2.6 – 1.9%, and 2.8 – 1.9% for nutrient solution A, nutrient solution B, and farmer practice treatments, respectively. Average tissue-N values were 2.51, 2.16, and 2.14% for the three respective treatments. The obtained values were less than reported for Azolla pinnata by Kushari and Watanabe (1991) with N (%) and by Shome (2024) with 4% tissue-N content. The low tissue-N content values obtained under the current experiment could be due to the high air temperatures that exceeded 35 oC for several months with direct effect on N-fixation by Azolla plants. The results agreed with those reported by Reddy (1987) who stated that maximum nitrogenase activity was observed between 20 – 30 oC and the rate of N₂ fixation was found to be highest at 30 oC.

As for tissue-P contents (%) in Azolla pinnata, results showed that average P values were 0.54, 0.40, and 0.40% for nutrient solution A, nutrient solution B, and farmer practice treatments, respectively. The high tissue-P values of 0.89, 0.62, and 0.61% recorded during December were due to the application of the 3rd dose of fertilizer by the end of November. The obtained results were close to those reported by Kushari and Watanabe (1991), who reported that P (%) in Azolla pinnata tissues varied from 0.40 to 1.04% as P-concentration in the growing media varied from 4.64 to 13.9 µg/cm²/day. Results were also close to those of Bhatt et al. (2020) who reported that mineral profiling of Azolla pinnata phosphorus was 0.31%. The obtained results were less than what was reported by Shome (2024), who conducted research on Azolla pinnata under open area conditions and showed that the plants had the capacity to accumulate a good quantity of tissue-P of 1.45% at higher concentration of media-P.

Regarding tissue-K contents (%), results indicated that tissue-K contents (%) in *Azolla pinnata* varied between 1.34 - 1.05%, 1.33 - 1.13%, and 1.4 - 1.14% for nutrient solution A, nutrient solution B, and farmer practice treatments, respectively. Average tissue-K values were 1.22, 1.21, and 1.25% for the three respective treatments. The obtained results were less than those reported for *Azolla pinnata* by Kushari and Watanabe (1991) with K (%) between 4.4 - 5.0%, Bhatt et al. (2020) with K-mineral profiling of 2.68%, and Shome (2024) with tissue-K of 4.30% in open area.

Water applied (WA) and its productivity (WP)

This experiment was the first in Egypt to measure and report the amounts of water needed to grow *Azolla* plants. Results in Table 6 showed that the total amount of applied water, including the effective rainfall during November, through the growing period (24 June - 31 December 2023) was 4071 m³/ha. The highest amount (1406 m³/ha) was added during July, while the lowest amount (281 m³/ha) was added during December. This trend is logical due to the fluctuation in the obtained *Azolla* yields and the prevailing weather condition at the experimental site during the growing period. The obtained results were different than those reported by Amro (2022) who reported that the monthly water consumption rate by *Azolla* plant was approximately equal to 120 m³/dunum (1200 m³/ha/month). The differences in the results could be due to the cultivated *Azolla* variety, fertilizers used and the prevailing weather conditions at the two sites.

Results indicated also that, monthly and total depths of water applied were much less than the corresponding ETo values (Table 6). The AW/ETo ratios varied from 0.23 during September to 0.44 during July with an average growing period value of 0.37. The results showed that, floating *Azolla* plays a significant role in decreasing evapotranspiration from the growing surface. The obtained results agreed well with those reported by Kimani et al. (2020) who reported that *Azolla* cover significantly decreased evapotranspiration (ET) losses compared with open water surfaces and green polyester covered mats. They concluded that the obtained results may be attributed to a greater total reflectance of the incoming solar radiation and enhanced modification of the surrounding microclimate by the dense mat of the floating *Azolla*. The obtained results were also close to those of Diara and Van Hove (1984) and Liu and Zheng (1992), who reported relative reduction in ET due to floating *Azolla* cover.

Results showed that, the highest average water productivity value (96.4 kg fresh yield/m³ water applied) was recorded for solution A treatment, while the lowest value (73.6 kg fresh yield/m³ water applied) was recorded for farmer practice treatment (Table 7). Also, the highest average WP value of 126.8 kg fresh yield/m³ water applied was obtained during October and the lowest value of 44.4 kg fresh yield/m³ water applied was obtained during July. The interaction effect of the tested nutrient solutions and the growing period showed that, the highest WP value of 141.3 kg fresh yield/m³ water applied was recorded for Solution A treatment during October, while the lowest value of 37 kg fresh yield/m³ water applied was recorded for farmer practice during July.

The initial amount of water added to fill the ponds (2249 m³/ha) was not included as water consumed because it can be reused easily for irrigating other crops in the farm. The chemical analysis of the ponded water by the end of the experiment is shown in Table 8. The chemical analysis indicated that the ponded water can provide essential crop nutrients.

CONCLUSION

Applying nutrient solution A that include multiple nutrients (i.e. Ca, K, P, Mg, B, Fe, Zn and Mn) significantly increased *Azolla pinnata* biomass and produced, on average, 54,9 t/ha/month.

Injecting the nutrients by vensuri to the ponded water was more effective than adding the nutrients by broadcasting or foliar spray.

Highest biomass production was obtained during summer.

Applied amount of water during the growing period, 24 June – 31 December 2023, was 4071 m³/ha.

An average water productivity value of 96.4 kg fresh biomass/m³ applied water was recorded for nutrient solution A.

The ponded water of 2249 m³/ha can be used to irrigate different crops in the farm.

REFERENCES

Abdul, Aziz (2012). New method of large-scale production of *Azolla pinnata* var. *pinnata* R. Brown: A Multipurpose crop. *International Journal of Applied Agricultural Research*, 7:1-9.

Adzman, N., Goh, S. J., Johari, A., Alam, M. Z., Kamaruddin, M. J. (2022). Preliminary study on *Azolla* cultivation and characterization for sustainable biomass source. *Journal of Physics: Conference Series*, 2259: 012018.

Alalade, O.A., E.A. Iyayi (2006). Chemical composition and the feeding value of *Azolla* (*Azolla pinnata*) meal for egg-type chicks. *International Journal of Poultry Science*, 5: 137-141.

Allen, R.G., L.S. Pereira, D. Raes, M. Smith. (1998). Crop evapotranspiration-Guidelines for computing crop water requirements. *FAO Irrigation and drainage paper* 56, FAO, Rome.

Amro, R.H.M. (2022). *Azolla* Cultivation on several types of water using several cheap natural fertilizers that are locally available in Palestine. M.Sc. Thesis. Al-Quds Univ. Palestine.

AOAC (2016). Official method of analysis, 20th edition, Association of Official Analytical Chemists, Washington, D.C. 5:137-141.

Ashraf, M.A., V.P. Sharma (2015). Role of *Azolla* in controlling mosquito breeding in Ghaziabad District Villages. *Indian J. Malariology*, 28: 51-54.

Beck, H.E., Zimmermann, N.E., McVicar, T.R., Vergopolan, N., Berg, A., Wood, E.F. (2018). Data descriptor: present and future Köppen-Geiger climate classification maps at 1-km resolution. *Scientific Data*, 5: 1-12.

Bhatt, N, R. Chandra, S. Kumar, K. Singh, N.P.S. Rajneesh, A.K. Singh, S.P.S. Chauhan (2020). Nutritive analysis of *Azolla pinnata* and its cultivation during winter season. *Int. J. Curr. Microbiol. App. Sci.*, 9: 2012-2018.

Biswas, M., Parveen, S., Shimozawa, H., Nakagoshi, N. (2005). Effects of *Azolla* species on weed emergence in a rice paddy ecosystem. *Weed Biology Management*, 5: 176-183.

Brouwer, P., H. Schluepmann, K.G. Nierop, J. Elderson, P.K. Bijl, I. van der Meer, W. de Visser, G-J. Reichart, S. Smeekeens, A. van der Werf. (2018). Growing *Azolla* to produce sustainable protein feed: the effect of differing species and CO₂ concentrations on biomass productivity and chemical composition. *Journal of the Science of Food and Agriculture*, 98: 4759-68.

Carithers, R.P., D.C. Yoch, and D.I. Arnon (1979). Two forms of nitrogenase from the photosynthetic bacterium *Rhodospirillum rubrum*. *J. Bacteriol.*, 137:779-789.

Chatterjee, A., P. Sharma, M.K. Ghosh, M. Mandal, P.K. Roy (2013). Utilization of *Azolla Microphylla* as Feed Supplement for Crossbred Cattle. *International Journal of Agriculture and*

Food Science Technology, 4: 207-214.

Chris, A., G. Luxmisha, J. Masih, G. Abraham (2011). Growth, photosynthetic pigments and antioxidant responses of *Azolla filiculoides* to monocrotophos toxicity. *Journal of Chemical and Pharmaceutical Research*, 3: 381-388.

Dastane, N.G. (1974). Effective rainfall in irrigated agriculture. F.A.O. Irrigation and Drainage Paper 25, FAO, Rome, Italy.

Diara, H.F., C. Van Hove. (1984). *Azolla*, a water saver in irrigated rice fields? In: Silver WS, Schröder EC (eds.) *Practical application of Azolla for rice production: Proceedings of an International Workshop*, Mayaguez, Puerto Rico, November 17-19, 1982. Springer Netherlands, Dordrecht, pp 115-118.

El-Naggar, Soad, H.S. El-Mesery (2022). *Azolla pinnata* as unconventional feeds for ruminant feeding. *Bulletin of the National Research Centre*, 2022: 46-66.

Hossain, M.A., S.A. Shimu, M.S.A. Sarker, M.E. Ahsan, M.R. Banu (2021). Biomass growth and composition of *Azolla* (*Azolla pinnata* R. BR.) supplemented with inorganic phosphorus in outdoor culture. *SAARC J. Agric.*, 19: 177-184.

Kannaiyan, S., M. Thangaraju, G. Oblisami (1981). Mineral nutrition of *Azolla*. *Tamil Nadu Agric. Univ. Newslett.*, 8: 2.

Kannaiyan, S. (1982). Mineral nutrition for *Azolla*. In: S. Kannaiyan and K. Kumar, editors, *Azolla biofertilizer for sustainable rice production*. Daya Publishing House, Delhi, India. p. 452.

Kavya, K. (2014). Nutritional evaluation of *Azolla* (*Azolla pinnata*) and its supplementary effect on in vitro digestibility of crop residues and total mixed ration. Ph.D. Thesis, Karnataka Veterinary, Animal and Fisheries Sciences University, Bidar.

Kimani, S.M., T. Kanno, K. Tawaraya, W. Cheng (2020). Floating *Azolla* cover influences evapotranspiration from flooded water surfaces. *Wetlands*, 40:1425-1432.

Kumar, M., T. Sharma, R.K. Dhuria (2018). A nutritional evaluation of *Azolla* (*Azolla pinnata*) as feed supplement. *Veterinary Practitioner*, 19: 132-133.

Kushari, D.P., I. Watanabe (1991). Differential responses of *azolla* to phosphorus deficiency I. Screening methods in quantity controlled-condition. *Soil Sci. Plant Nutr.*, 37: 271-282.

Liu, C.C., W.W. Zheng (1992). Nitrogen fixation of *Azolla* and its utilization in agriculture in China. In: Hong G.F (Ed). *The nitrogen fixation and its research in China*. Springer Berlin Heidelberg, Berlin, Heidelberg. pp: 525-537.

Mohamed, M., S. Elnemir, S. Abd El-Mounem, S. Abo El-Maati (2018). *Azolla* fern as untraditional resource of protein. *Zagazig J. Agric. Res.*, 45:1345-1355.

Mohan, K.R., B. Patil, Yamanura (2020). *Azolla*: The wonder fern. *Krishi Sci. eMagazine for Agricultural Science*, p. 25-27.

Mosha, S.S.A. (2018). Review on significance of *Azolla* meal as a protein plant source in finfish culture. *J. Aquac. Res. Dev.*, 9:1000544.

Nordiah, B., Z. Muta Harah, B. Japar Sidik, W.N. Wan Hazma. (2012). *Azolla pinnata* growth performance in different water resources. *Pakistan J. of Bio. Sci.*, 15: 621-628.

- O'Hara, G.W. (2001). Nutritional constraints on root nodule bacteria affecting symbiotic nitrogen fixation: a review. *Aust. J. Exp. Agric.*, 41: 417-433.
- Pabby, A., R. Prasanna, P.K. Singh (2003). Azolla-Anabena symbiosis from traditional agriculture to Biotechnology. *Indian Journal of Biotechnology*, 2: 26-37.
- Patil, P.V., M.K. Patil (2020). *Milk Production Management*. London: CRC Press.
- Rashad, S. (2021). An overview on the aquatic fern *Azolla* spp. as a sustainable source of nutrients and bioactive compounds with resourceful applications. *Egyptian Journal of Aquatic Biology & Fisheries*, 25: 775-782.
- Ray, T.B, B.C. Mayne, R.E. Toia, G.A. Peters (1979). Azolla-Anabaena relationship. VIII. Photosynthetic characterization of the association and individual partners. *Plant Physiology*, 64: 791-795.
- Reddy, K.R. (1987). Nitrogen fixation by *Azolla* cultured in nutrient enriched waters. *J. Aquat. Plant Manage.*, 25: 43-48.
- Sadeghi, R., R. Zarkami, K. Sabetraftar, P. Van Damme (2012a). Use of support vector machines (SVMs) to predict distribution of an invasive water fern *Azolla filiculoides* (Lam.) in Anzali wetland, southern Caspian Sea, Iran. *Ecological Modelling*, 244: 117-126.
- Sadeghi, R., R. Zarkami, K. Sabetraftar, P. Van Damme (2012b). Application of classification trees to model the distribution pattern of a new exotic species *Azolla filiculoides* (Lam.) at Selkeh Wildlife Refuge, Anzali wetland, Iran. *Ecological Modelling*, 243: 8-17.
- Sadeghi, R., R. Zarkami, K. Sabetraftar, P. Van Damme (2013). A review of some ecological factors affecting the growth of *Azolla* spp. *Caspian J. Env. Sci.*, 11: 65-76.
- Selvaraj, K., R. Chowdhury, C. Bhattacharjee (2014). Optimization of the solvent extraction of bioactive polyphenolic compounds from aquatic fern *Azolla microphylla* using response surface methodology. *Int. Food Res. J.*, 21:1559-1567.
- Shome, G. (2024). The potential capabilities of two species of *Azolla* as a bioaccumulator in summer. *Environment and Ecology*, 42: 828-835.
- Snedecor, G.W., W.G. Cochran (1981). *Statistical methods*, 6th edition. Ames, IA: Iowa State University Press, USA.
- Subudhi, B., P. Singh (1978). Nutritive value of the water fern *Azolla pinnata* for chicks. *Poultry Science*, 57: 378-380.
- Sujatha, Kundu, Jeyakumar, M. Kundu. (2013). Azolla supplementation: Feed cost benefit in duck ration in Andaman Islands. *Tamil Nadu J. Vet. and Ani Sci.*, 9: 130-136.
- Tan, K.H. (1996). *Soil sampling, preparation and analysis*. New York, NY: Marcel Dekker
- Brockhaus, F. A. 1962. *A B C der landwirtschaft B. and A-K* 2nd Edit VEB F. A. Brockhaus Verlag, Leipzig.
- VanHove, C., Y. Lopez (1987). Fisiologia de *Azolla*. In: Preston, T.R. and E. Murgueitio. *Trees and shrub Workshop on the assessment of Azolla use in legumes as protein sources for livestock*. In: *Forage tropical Latin America*. Chicklayo: Peru.
- Wagh, H.A., C.S. Desai, M.N. Ambhore, P.S. Kamble P.D. Kamble (2021). Production of azolla in

different condition and its comparative study. *International Journal of Advanced Biotechnology and Research*, 12: 22-25.

Wagner, G.M. (1997). Azolla: A review of its biology and utilization. *The Botanical Review*. 63: 1-26.

Watanabe, I., C.R. Espinas, N.S. Berja, B.V. Alimagno (1977). Utilization of Azolla-Anabaena complex as a nitrogen fertilizer for rice. *Intl. Rice Res. Inst. Res. Pap. Ser.*, 11: 1-15.

Yee, T.J., N.A. Kamaruddin, S.S. Safiah Mohamad. (2022). Nutritional evaluation of Azolla pinnata and Azolla microphylla as feed supplements for dairy ruminants. *Journal of Agrobiotechnology*, 13: 17-23.

Acknowledgements

Authors want to acknowledge the full support of Prof. Drs. Mohamed El-Khouly; Director of SWERI, Sayed El-Tohamy; Head of the Central Laboratory at SWERI, and Nader Habashy; member of the Central Lab. at SWERI for providing all research facilities including moral and technical support during the study. The authors which to express their deep thanks to Mr. Moataz Shafik, Mr. Asser Gouname, Mr. Hatem Magdy, and Mr. Mohamed Mosharbak for supporting the researchers while conducting this research at their private farm.

References